Compounds of unstable DP IV-inhibitors

The present invention relates to compounds of unstable
inhibitors of dipeptidyl peptidase IV (DP IV), which
compounds have the general formula A-B-C, wherein

A is an amino acid,

B is a chemical bond between A and C or is an amino acid,
and

C is an unstable inhibitor of DP IV.

Aminoacyl thiazolidides, aminoacyl pyrrolidides, N-dipep-
tididyl, O-acyl hydroxylamines and other compounds are
known to act as inhibitors of the serum enzyme dipeptidyl
peptidase IV (DP IV) and of analogous enzymes [see DEMUTH,
H.-U., J. Enzyme Inhibition 3, 249 (1990); DEMUTH, H.-U.,
HEINS, J., in *Dipeptidyl Peptidase IV* (B. Fleischer, Ed.)
R.G. Landes, Biomedical Publishers, Georgetown, 1995, 1-
37].

It has been found that the T-cell-mediated immune response,
for example in the case of transplantations, is influenced
by means of stable inhibitors of dipeptidyl peptidase IV,.
[see KOROM, S., DEMEESTER, I., STADLBAUER, T.H.W.,
CHANDRAKER, A., SCHAUB, M., SAYEGH, M.H., BELYAEV, A.,
HAEMERS, A., SCHARPE, S., KUPIEC-WEGLINSKI, J.W.,
Inhibition of CD26/dipeptidyl peptidase IV activity in vivo
prolongs cardiac allograft survival in rat recipients,
Transplantation 63, 1495 (1997)]. Rheumatoid arthritis can
also be suppressed [see TANAKA, S., MURAKAMI, T., HORIKAWA,
H., SUGIURA, M., KAWASHIMA, K., SUGITA, T., Suppression of

arthritis by the inhibitors of dipeptidyl peptidase IV.
Int. J. Immunopharmacol. 19, 15 (1997)].

It has also been found that, because of the associated
temporary reduction in enzyme activity, administering
stable inhibitors (effectors) of DP IV or of DP IV-
analogous enzyme activity in the blood of a mammal causes
reduced breakdown of the endogenous (or additionally
exogenously administered) insulinotropic peptides gastric
inhibitory polypeptides 1-42 (GIP$_{1-42}$) and glucagon-like
peptide amides-1 7-36 (GLP-1$_{7-36}$) (or GLP-1$_{7-37}$ or analogues
thereof) by DP IV and DP IV-like enzymes and, accordingly,
the drop in concentration of those peptide hormones or
analogues thereof is reduced or delayed. The greater
stability of the (endogenously present or exogenously
introduced) incretins or analogues thereof, brought about
by the action of the DP IV-effectors, which increases their
availability for insulinotropic stimulation of the incretin
receptors of the Langerhans cells in the pancreas, *inter*
alia alters the effectiveness of the body's own insulin,
resulting in stimulation of carbohydrate metabolism in the
organism treated. As a result, the blood sugar level in
the serum of the organism treated falls below the glucose
concentration characteristic of hyperglycaemia.
Consequently, by means of DP IV-inhibitors it is possible
to prevent or alleviate metabolic anomalies, such as excess
weight, glucosuria, hyperlipidaemia, and possible serious
metabolic acidoses and diabetes mellitus, which are the
result of prolonged elevated glucose concentrations in the
blood [see DE 196 16 486].

By means of DP IV-inhibitors it is also possible, experimentally, to prevent the penetration of CD 26 (DP IV) positive cells by HIV [see WAKSELMAN, M., NGUYEN, C., MAZALEYRAT, J.-P., CALLEBAUT, C., KRUST, B., HOVANESSIAN, A.G., Inhibition of HIV-1 infection of CD 26+ but not CD26- cells by a potent cyclopeptidic inhibitor of the DPP IV activity of CD 26. Abstract P 44 of the 24th European Peptide Symposium 1996].

It has also been found that DP IV can modulate the activity of neuroactive peptides, such as neuropeptide Y and CLIP [see MENTLEIN, R., DAHMS, P., GRANDT, D., KRUGER, R., Proteolytic processing of neuropeptide Y and peptide YY by dipeptidyl peptidase IV. Regul. Pept. 49, 133 (1993); WETZL, W., WAGNER, T., VOGEL, D., DEMUTH, H.-U., BALSCHUN, D., Effects of the CLIP fragment ACTH 20-24 on the duration of REM sleep episodes. Neuropeptides, 31, 41 (1997)].

These varied actions of DP IV-inhibitors suggest that, when used in the case of a specific pathophysiological condition of a tissue, their actions can have an effect on other normal physiological states, for example in other organs. Those effects can have both positive and negative consequences for the target organism.

The problem of the present invention is accordingly to provide effectors of DP IV that have a high bioavailability of DP IV-inhibitors and a precisely defined duration of action in specific target tissues or target organs.

It was especially a problem of the present invention to provide inhibitors of DP IV that have a precisely defined

short duration of action combined with high bio-
availability.

Those problems are solved by the provision of compounds of
unstable inhibitors of dipeptidyl peptidase IV (DP IV),
which compounds have the general formula A-B-C, wherein

A is an amino acid,
B is a chemical bond between A and C or is an amino acid,
and
C is an unstable inhibitor of DP IV.

When B represents a bond, it is especially a peptide bond;
when B is an amino acid, it is preferably linked to A and C
by way of peptide bonds.

According to the invention, those compounds can be used as
inhibitors of DP IV, it being possible for the site of
their action, the time of onset of their action and the
duration of action to be defined precisely:

Upon administration, the compounds are cleaved, for example
by suitable enzymes, and the unstable "C" inhibitors are
thus released by removal of the groups A-B. The inhibitors
are released both by chemical and enzymatic mechanisms.
For example, esterases, proteases and peptidases serve to
release the active ingredient from compounds according to
the invention. Such esterases, proteases, etc., are
disclosed, for example, in WO 97/45117, US 5433955,
US 5614379 and US 5624894.

The released unstable inhibitors can then interact with the
DP IV already present and inhibit it. As a direct result,
for example, the above-mentioned insulinotropic peptides
are broken down to a lesser degree and the effectiveness of
insulin is thereby increased.

The administration of unstable inhibitors of DP IV *per se*
has proved disadvantageous for inhibiting DP IV since they
are broken down very rapidly *in vivo* and thus uniform
distribution of the inhibitors, especially in the human
body, is impossible. In particular, upon oral
administration such inhibitors are so unstable that they
have virtually no activity at all. Accordingly, stable
inhibitors have hitherto been used especially in the
treatment of diabetes mellitus.

Surprisingly it has now been found that unstable "C"
inhibitors can be stabilised sufficiently by masking them
as groups of the formula A-B-C.

That stabilisation is also surprising insofar as a compound
of formula A-B-C that has a carbonylmethyl pyridinium group
is positively charged at the pyridinium nitrogen atom. As
a result, electron-attraction is exerted by the methylene
group upon the group that, after removal of the A-B
radical, constitutes the active nucleophilic reaction
centre of the inhibitor. On the basis of the resulting
activation of the reaction centre, it would have been
expected that its nucleophilic tendency would have been
increased to such an extent that nucleophiles would "dock"
non-specifically at the compound A-B-C and the inhibitor
would be inactivated. Surprisingly it has been found,

however, that such inactivation of the inhibitors does not occur.

In order to intervene by means of DP IV-inhibitors in physiological control circuits that require only a short-term effect upon the target enzyme DP IV, according to the invention there are provided, for example, inhibitors as component C that have only a short duration of action and that after a definable half-life become chemical compounds that have no inhibitory activity.

For example, to strengthen the incretin effect in diabetes mellitus a duration of action of the inhibitors of a few minutes is sufficient whilst, for example, the suppression of the DP IV-mediated immune response in the case of transplantations requires long-term action by the inhibitors.

After being released, the unstable inhibitors according to the invention cyclise, for example, into a piperazine derivative and are thus inactivated. That reaction occurs spontaneously and is attributable to the nucleophilic action of the N-terminal amino nitrogen on the C-terminal carbonyl function of the dipeptide derivative and is facilitated by *cis/trans* isomerisation around the amino acid/imide bond, which isomerisation is facilitated especially in proline-containing peptides.

Moreover, that decomposition process does not begin until the compound has reached the desired target compartment, for example the blood circulation, and has begun the desired activity.

Those properties of the inhibitors according to the
invention can be used according to the invention to design
different DP IV-inhibitors in order to trigger the desired
temporally defined deactivation of the DP IV-inhibitor by
intramolecular cyclisation after it has been released.

In particular, according to the invention preference is
given to compounds in which C is a dipeptide derivative
having an active carbonyl group at the C-terminus.
Preferably C is a dipeptidyl chloroalkyl ketone, dipeptidyl
boronic acid or dipeptidyl cyanide compound or a dipeptidyl
pyridinium methyl keto compound. Such inhibitors have
proved to be especially effective unstable DP IV-
inhibitors. There may be mentioned as examples of the
dipeptide group, for example Ile-Thia, Ile-Pyr, Val-Thia
and Val-Pyr.

According to the invention, the inhibitors (component C)
may also be present in salt form, with preference being
given to organic salts, such as acetates, succinates,
tartrates or fumarates, or to inorganic acid radicals, such
as phosphates or sulphates. Special preference is given to
fumarates (?).

According to a preferred embodiment of the present
invention, compounds are used in which B is proline,
hydroxyproline, thiazolidinecarboxylic acid, dehydro-
proline, pipecolic acid, azetidinecarboxylic acid or
aziridinecarboxylic acid, with proline and hydroxyproline
being especially preferred.

In particular, the compounds according to the invention also have the advantage that the inhibitors of DP IV are released according to individual patients' needs:

When a compound according to the invention interacts with a DP IV molecule, it is cleaved by the enzyme into the groups A-B and the inhibitor C. The inhibitor C will inhibit the DP IV molecule so that it cannot cleave any further compounds. If further DP IV molecules are present, the compounds will continue to be cleaved (if a sufficient amount of corresponding compounds has been administered) until the last DP IV molecule has been inhibited. The remaining compounds are not broken down and thus constitute an inhibitor reservoir until the concentration of DP IV molecules rises again or inhibitor molecules are displaced by the DP IV or inhibitor molecules are eliminated or inactivated, and the compounds according to the invention are then cleaved again, thus releasing inhibitors.

The invention also has the further advantage that each organism will release the exact amount of inhibitor that is necessary to inhibit the amount of DP IV present, which is different in individual cases. If, for example, a patient has a high concentration of DP IV then a large amount of inhibitor will be released; if there is only a slightly elevated concentration of DP IV, only a small amount of inhibitor will be released.

In particular, special preference is given to compounds in which A-B is a dipeptide of the formula Ile-Pro or Gly-Pro (?).

The present invention accordingly relates to novel compounds of unstable inhibitors of the serine peptidase dipeptidyl peptidase IV, which compounds can be used in the treatment of various disorders, especially of metabolic disorders associated with diabetes mellitus.

Surprisingly such masked inhibitors are additionally considerably more effective than non-masked inhibitors: if identical amounts of non-masked DP IV-inhibitors and of compounds according to the invention are used, the compounds according to the invention produce a marked improvement in glucose tolerance in Wistar rats (?).

A further advantage of the compounds according to the invention lies in the fact that the onset of action and also the duration of action of the DP IV-inhibitors can be temporally controlled by suitable selection of the groups A-B. In particular, the release of the groups A-B from the compounds according to the invention depends upon the nature of the amino acid radical of A: in respect of the definition of group A, the following sequence has been found in particular for the rate at which the radicals A-B are released from the compounds A-B-C by DP IV: Ile<Val<Phe<Pro<Ala<Gly. The rate constants of the corresponding DP IV-catalysed release are from 1 s^{-1} to 100 s^{-1}. A means is thus available for releasing the DP IV-inhibitors in a precisely temporally defined manner: if the enzymes are to act immediately, for example upon intake of glucose-rich nutrient, a compound A-B-C will be selected that has, for example, the amino acid Gly as the A group; if the action of the inhibitor is to be delayed then the amino acid Ile, for example, can be selected as group A.

By means of the compounds according to the invention, it is thus possible for the DP IV-inhibitors to be transported through the mucosa of the small intestine especially virtually without delay, for example virtually simultaneously with nutrient intake.

Moreover, the site at which the DP IV-inhibitors are released and at which they act can also be controlled by the nature of the radicals A-B:

Various other amino peptidases, such as, for example, pyroglutamyl aminopeptidase and prolyl aminopeptidase, are present in the blood of mammals in addition to dipeptidyl peptidase IV. By suitable selection of the radicals A-B, it is possible according to the invention to determine the aminopeptidase by which the DP IV-inhibitor is to be released and so to determine where the action of the inhibitor is to occur. The compounds according to the invention or corresponding pharmaceutical compositions can thus also be used in cell-, tissue- or organ-specific inhibition of DP IV. The groups A-B can also be so selected that only those enzymes that are present only vascularly and that release the inhibitors at a sufficiently fast rate are targeted.

To summarise, it may be stated that by means of the compounds of unstable DP IV-inhibitors according to the invention, it is possible, in a completely surprising manner:

1. to achieve increased action of the inhibitors;

2. for the inhibitors to be released according to patients' needs;

3. for the inhibitors to be released from the compounds in a temporally controlled manner;

4. for the site at which the inhibitors are released from the compounds to be controlled;

5. for a reservoir of DP IV-inhibitors to be provided; and

6. for the duration of action or the end of action of the initiators to be precisely defined from the time of their unmasking.

According to the invention there are also provided pharmaceutical compositions especially for oral administration, characterised in that they comprise at least one compound according to the invention optionally in combination with customary carriers or excipients.

The compounds according to the invention or pharmaceutical compositions comprising them can be used in the treatment or prophylaxis of disorders in mammals that can be treated by modulating the DP IV activity of a mammal, such as, for example, metabolic disorders in humans.

In particular they can be used in the treatment of impaired glucose tolerance, glucosuria, hyperlipidaemia, metabolic acidoses, diabetes mellitus, diabetic neuropathy and nephropathy and sequelae of diabetes mellitus in mammals.

Because of the preferably short duration of action of the
unstable inhibitors according to the invention, it is
especially possible to minimise or prevent an influence
upon processes in the human or animal body that would
require long-term inhibition of DP IV.

Embodiments

**Synthesis of unstable DP IV-inhibitors C and compounds of
unstable DP IV-inhibitors according to the invention
(A-B-C)**

The preparation of unstable DP IV-inhibitors C
(corresponding to the general formula A-B-C) is shown in
Examples 1.1 and 1.2. The synthesis of a compound of
unstable DP IV-inhibitors according to the invention is
shown in Example 1.3. The starting materials, the
corresponding peptidyl chloromethyl ketones in each case,
were prepared according to known processes (WEINSTEIN, B.,
Chemistry and Biochemistry of Amino Acids, Peptides, and
Proteins, Marcel Dekker, New York, Basle, 1977). The
pyridinium methyl ketones prepared by way of example
under 1.1. and 1.2, which are N-terminal-protected
dipeptide derivatives, are extremely stable and can be
characterised completely. At normal air humidity, the N-
terminal deblocked dipeptide derivatives begin the
intramolecular decomposition process immediately after
being deblocked, so that it is not possible to determine a
melting point. The products are characterised by means of
HPLC and mass spectrometry.

1.1 Synthesis of H-Val-Pro-CH₂-(N⁺C₅H₅)/Cl⁻

a) Z-Val-Pro-CH₂-(N⁺C₅H₅)/Cl⁻

Structural formula:
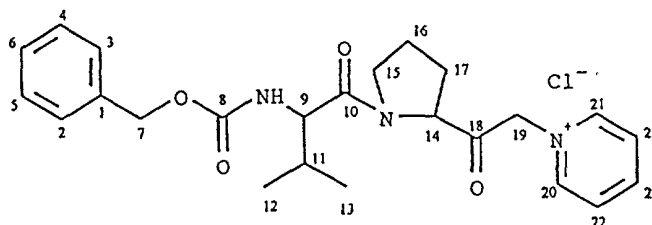
Preparation:	10 mmol of Z-Val-Pro-chloromethyl ketone are dissolved in pyridine and stirred at 25°C for 5 days. The excess pyridine is distilled off at 2 mbar vacuum. The Z-Val-Pro-pyridinium methyl ketone is subjected to HPLC purification. The compound is an oil.

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Empirical formula:	$C_{24}H_{30}N_3O_4Cl$
Molecular weight:	459.97 Da
Yield:	45.8 % of the theory
HPLC:	retention time: 2.3 min, LiChrosper 100 RP-18 (125-4), λ = 220 nm, flow rate 0.5 ml/min, isocratic 50% acetonitile in H_2O) (0.1 % TFA) retention time: 19.3 min, Nucleosil 7 C_8, λ = 220 nm, flow rate 8 ml/min,

isocratic 50 % acetonitrile in H_2O

(0.1 % TFA)

^{1}H NMR (DMSO-d_6) δ_H: 0.8-1.0 (6H, m, H_{12} and H_{13}), 1.8-2.1 (3H, m, H_{11} and H_{16}), 2.2-2.4 (2H, m, H_{17}), 3.4-3.7 (2H, m, H_{15}), 3.7-4.1 (2H, m, H_{20} and H_{21}), 4.3-4.5 (1H, dd, 5 Hz, 8 Hz, H_9), 4.8-5.1 (2H, m, H_7), 5.8-6.2 (3H, m, H_{14}, H_{19}), 7.2-7.5 (5H, m, H_2-H_6), 8.2-8.3 (2H, m, H_{22} and H_{23}), 8.6-8.7 (1H, m, H_{24}), 8.8-9.0 (1H, d, 6 Hz, NH)

^{13}C NMR (DMSO-d_6) δ_C: 136.8 (C_1), 127.9 (C_2, C_3), 127.8 (C_4, C_5), 128.4 (C_6), 66.7 (C_7), 156.3 (C_8), 57.9 (C_9), 170.8 (C_{10}), 29.7 (C_{11}), 18.5 (C_{12}, C_{13}), 63.7 (C_{14}), 47.2 (C_{15}), 25.1 (C_{16}), 27.9 (C_{17}), 200.3 (C_{18}), 71.1. (C_{19}), 146.1 (C_{20}, C_{21}), 128.3 (C_{22}, C_{23}), 146.4 (C_{24})

MALDI-TOF-MS m/z: 424.6 Da (M+H$^+$, without chloride anion)

b) H-Val-Pro-CH_2-($N^+C_5H_5$)/Cl$^-$

Structural formula:
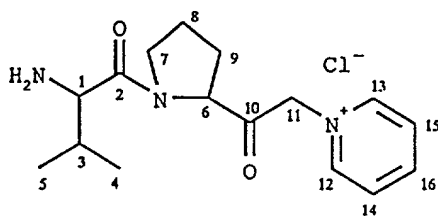
Preparation: The Z-protecting group is removed from Z-Val-Pro-CH$_2$-(N$^+$C$_5$H$_4$)/Cl$^-$ by means of HBr/glacial acetic acid in a reaction time of 5 minutes. 2 ml of HBr/glacial acetic acid (33%) are added to 1.0 mmol of Z-protected peptide and the mixture is stirred at 23°C for approx. 10 min. Concentration *in vacuo* is then carried out. The peptide is precipitated in the form of the hydrobromide from methanol by means of diethyl ether, suction-filtered and dried *in vacuo*.

Empirical formula: C$_{16}$H$_{24}$N$_3$O$_2$Cl

Molecular weight: 325.84 Da
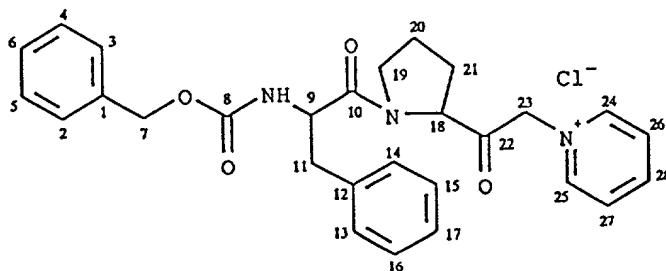
Yield: 97.7 % of the theory

HPLC: retention time: 7.4 min, LiChrosper 100 RP-18 (125-4), λ = 220 nm, flow rate 0.5 ml/min, isocratic 50% acetonitrile in H$_2$O (0.1% TFA)

MALDI-TOF-MS m/z: 291.2 Da (M+H$^+$, without chloride anion)

1.2   **Synthesis of H-Phe-Pro-CH$_2$-(N$^+$C$_5$H$_5$)/Cl$^-$**

a) Z-Phe-Pro-CH$_2$-(N$^+$C$_5$H$_5$)/Cl$^-$

Structural formula:

Preparation:	2 ml of pyridine are added to 10 mmol of Z-Phe-Pro-chloromethyl ketone. The mixture is stirred at 23°C for 4 days. The excess pyridine is distilled off at 2 mbar vacuum. The crude product is purified over 60 g of silica gel. In the chloroform/methanol eluate (9:1 parts by volume) initially the product is collected and, as the polarity of the eluant increases, the chloromethyl ketone is collected. Z-Phe-Pro-pyridinium methyl ketone is finally subjected to HPLC purification.

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Empirical formula:	$C_{28}H_{30}N_3O_4Cl$
Molecular weight:	508.01 Da
Yield:	69.6 % of the theory
HPLC:	retention time: 17 min, LiChrosorb RP 8 (Hibar), λ = 220 nm, flow rate 8.0 ml/min, isocratic 50 % acetonitrile in H_2O (0.1 % TFA)
	retention time: 3.4 min, LiChrosper RP-8 (125*4), λ = 220 nm, flow rate 1.5 ml/min, gradient 30-80% acetonitrile in H_2O (0.1 % TFA) in 25 min
	retention time: 10.2 min, Nucleosil 7 C$_8$, λ = 220 nm, flow rate 8 ml/min,

isocratic 50% acetonitrile in H_2O (0.1 % TFA)

^{1}H NMR (DMSO-d_6) δ_H: 1.7-2.1 (4H, m, H_{20}, H_{21}), 2.7-3.0 (2H, m, H_{11}), 3.4-3.9 (2H, m, H_{19}), 4.4-4.6 (1H, m, H_9), 4.6-4.8 (2H, m, H_{24}, H_{25}), 5.0-5.1 (2H, dd, H_7), 5.7-5.8 (1H, d, H_{18}), 5.9-6.1 (2H, dd, H_{23}), 7.2-7.4 (10H, m, H_2-H_6, H_{13}-H_{17}), 8.6-8.8 (1H, dd, H_{28}), 8.2-8.3 (2H, d, H_{26} and H_{27}), 8.8-8.9 (1H, d, NH)

^{13}C NMR (DMSO-d_6) δ_C: 136.8 (C_1), 127.9 (C_2, C_3), 127.6 (C_4, C_5), 128.4 (C_6), 65.5 (C_7), 156.3 (C_8), 54.1 (C_9), 170.9 (C_{10}), 36.4 (C_{11}), 137.6 (C_{12}), 126.6 (C_{13}, C_{14}), 128.4 (C_{15}, C_{16}), 129.3 (C_{17}), 63.1 (C_{18}), 46.9 (C_{19}), 25.1 (C_{20}), 27.6 (C_{21}), 200.7 (C_{22}), 66.3 (C_{23}), 146.2 (C_{24}, C_{25}), 128.2 (C_{26}, C_{27}), 146.4 (C_{28})

MALDI-TOF-MS m/z: 472.8 Da (M+H$^+$, without chloride anion)

b) H-Phe-Pro-CH_2-(N$^+C_5H_5$)/Cl$^-$
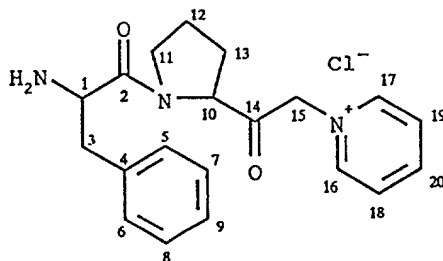
Structural formula:

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Preparation:	The Z-protecting group is removed from Z-Phe-Pro-CH$_2$-(N$^+$C$_5$H$_4$)/Cl$^-$ after a reaction time of 5 minutes. 2 ml of HBr/glacial acetic acid (33%) are added to 1.0 mmol of Z-protected peptide and the mixture is stirred at 23°C for approx. 10 minutes. Concentration *in vacuo* is then carried out. The peptide is precipitated in the form of the hydrobromide using diethyl ether, suction-filtered and dried *in vacuo*.
Empirical formula:	C$_{20}$H$_{24}$N$_3$O$_2$Cl
Molecular weight:	373.88 Da
Yield:	98 % of the theory
HPLC:	retention time: 6.9 min, LiChrosper 100 RP-18 (125-4), λ = 220 nm, flow rate 0.5 ml/min, isocratic 50% acetonitrile in H$_2$O (0.1 % TFA)
MALDI-TOF-MS m/z:	337.2 Da (M+H$^+$, without chloride anion)

1.3   Synthesis of H-Gly-Pro-Val-Pro-CH$_2$-(N$^+$C$_5$H$_5$)/Cl$^-$

a) Z-Gly-Pro-Val-Pro-CH$_2$-(N$^+$C$_5$H$_5$)/Cl$^-$

Structural formula:
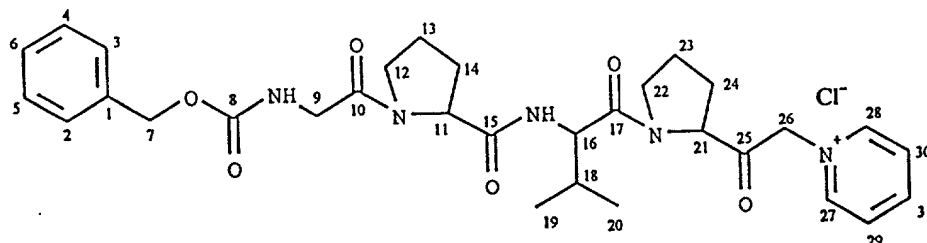
Preparation:	2 ml of pyridine are added to 10 mmol of Z-Gly-Pro-Val-Pro-chloromethyl ketone. The mixture is stirred at 23°C for 4 days. The excess pyridine is distilled off at 2 mbar vacuum. Z-Gly-Pro-Val-Pro-pyridinium methyl ketone is subjected to HPLC purification.
Empirical formula:	$C_{31}H_{40}N_5O_6Cl$
Molecular weight:	614.14 Da
HPLC:	retention time: 17.4 min, LiChrosorb RP 8 Hibar, $\lambda = 220$ nm, flow rate 8 ml/min, isocratic 50% acetonitrile in H_2O (0.1 % TFA)
	retention time: 5.4 min, LiChroCART 100 RP-18 (250-4), $\lambda = 220$ nm, flow rate 0.5 ml/min, isocratic 50% acetonitrile in H_2O (0.1 % TFA)
	retention time: 17.7 min, Nucleosil 100 7 C_8, $\lambda = 220$ nm, flow rate 5 ml/min, isocratic 50% acetonitrile in H_2O (0.1 % TFA)

^{13}C NMR (DMSO-d_6) δ_C: 134.4 (C_1), 128.9 (C_2, C_3), 128.2 (C_4, C_5), 129.8 (C_6), 65.3 (C_7), 157.2 (C_8), 39.0 (C_9), 165.7 (C_{10}), 56.0 (C_{11}), 41.6 (C_{12}), 24.6 (C_{13}), 29.0 (C_{14}), 170.5 (C_{15}), 52.1 (C_{16}), 171.9 (C_{17}), 30.3 (C_{18}), 18.6, 19.3 (C_{19}, C_{20}), 58.9 (C_{21}), 47.2 (C_{22}), 25.0 (C_{23}), 29.4 (C_{24}), 196.5

(C25), 65.8 (C26), 137.9 (C27, C28), 129.1 (C29, C30), 146.5 (C31)

MALDI-TOF-MS m/z: 579.7 Da (M+H$^+$, without chloride anion)

b) H-Gly-Pro-Val-Pro-CH2-(N$^+$C5H5)/Cl$^-$
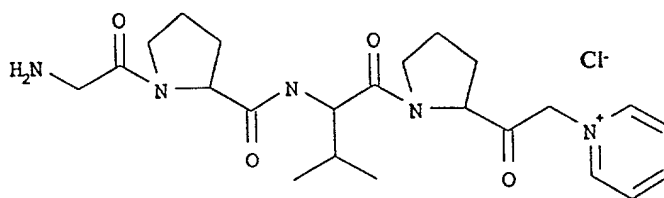
Structural formula:

Preparation:	The Z-protecting group is removed from Z-Phe-Pro-CH2-(N$^+$C5H4)/Cl$^-$ after a reaction time of 5 minutes. 2 ml of HBr/glacial acetic acid (33%) are added to 1.0 mmol of Z-protected peptide and the mixture is stirred at 23°C for approx. 10 minutes. Concentration *in vacuo* is then carried out. The peptide is precipitated in the form of the hydrobromide using diethyl ether, suction-filtered and dried *in vacuo*.
Empirical formula:	C23H34N5O4Cl
Molecular weight:	480.0 Da
Yield:	95 % of the theory
MALDI-TOF-MS m/z:	443.9 Da (M+H$^+$, without chloride anion)

1. Breakdown of unstable DP IV-inhibitors and of their masked forms in aqueous solution

To analyse the stability of the inhibitors prepared under
1.1 and 1.2, the inhibitors were incubated in an aqueous
buffer solution and their intramolecular cyclisation
reaction was monitored by means of MALDI-TOF mass
spectrometry (Figures 2 and 3). The products of that
reaction are the respective pyrazine derivatives
(Figure 1).

The breakdown of H-Phe-Pro-pyridinium metyl ketone having
the molecular weight of 337.2 Da to the cyclic pyrazine
derivative having the molecular weight of 319.2 Da, with
removal of water, is completed quantitatively within a
period of 30 minutes (Figure 2).

The breakdown of H-Val-Pro-pyridinium methyl ketone having
the molecular weight of 291.2 Da to the cyclic pyrazine
derivative having the molecular weight of 273.2 Da, with
removal of water, is completed quantitatively within a
period of 60 minutes (Figure 3).

The formation of the double bond system of the pyrazine,
which takes place during the intramolecular reaction,
enables quantitative analysis of the cyclisation process by
means of UV spectrometry (Figure 4). The rate constants
determined therefrom for the intramolecular cyclisation of
unstable DP IV-inhibitors in 0.1M HEPES buffer, pH = 7.6,
25°C, are shown in Table 1.

Table 1: Parameters of the cyclisation of unstable DP IV-
inhibitors

Compound	k min^{-1}	Half life (min)
H-Val-Pro-CH$_2$-(N$^+$C$_5$H$_5$)	8.7 x 10^{-4} (± 4x10^{-5})	13.3
H-Phe-Pro-CH$_2$-(N$^+$C$_5$H$_5$)	1.2 x 10^{-3} (± 3.9x10^{-5})	9.6

In contrast thereto, the compounds H-Val-Pro-CH$_2$-(N$^+$C$_5$H$_5$) and
H-Gly-Pro-Val-Pro-CH$_2$-(N$^+$C$_5$H$_5$) according to the invention
have proved to be completely stable over a period of
24 hours under identical conditions.

**2.   Interaction of unstable DP IV-inhibitors or compounds
comprising dipeptidyl peptidase IV according to the
invention in aqueous solution**

When the DP IV target enzyme is incubated with unstable
inhibitors in the presence of a substrate, inhibition of
the enzyme is observed initially; the inhibition subsides
again as the experimental time progresses as a result of
the intramolecular cyclisation of the inhibitor which takes
place in parallel, since the concentration of the inhibitor
in the reaction solution decreases as a result of the
spontaneously occurring chemical reaction. That effect is
shown in Figures 5 and 6. Because of the time-dependent
drop in the concentration of the inhibitor, the rate of the
enzyme-catalysed hydrolysis of the substrate increases
again as time progresses.

In contrast to the non-masked DP IV-inhibitors, the compound H-Gly-Pro-Val-Pro-CH_2-($N^+C_5H_5$) according to the invention has proved to be stable over a period of 24 hours in buffered aqueous solution in the absence of an enzyme. The active DP IV-inhibitor H-Val-Pro-CH_2-($N^+C_5H_5$) is released only as a result of the addition of the DP IV enzyme (used here by way of example also to release the DP IV-inhibitor) with removal of the N-terminal dipeptide H-Gly-Pro-OH. Accordingly, in the mass spectrum (Figure 7) markedly more than 50% of the incubated compound according to the invention can be detected even after an incubation period of 60 minutes. Owing to that delayed release, surprisingly there is also observed, in addition to the desired effective inhibition of the target enzyme, a markedly prolonged activity combined with markedly reduced concentration of the compound according to the invention compared with the unstable DP IV-inhibitors (Figure 8).